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DETERMINATION OF AMILORIDE VIA QUENCHED CONTINUOUS FLUORESCENCE OF AZO DYE USING LOW-PRESSURE MERCURY LAMP TUBE (UV-LIGHT) AND MULTI SOLAR CELLS AT 2 X 90° AS A DETECTORS

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Abstract

Purpose: The purpose of this approach was to provide a new, easy, and fast way to use fluorescence measurements to determine the concentration of pharmaceutical medicines and Amiloride (AMD) in their pure form.

Keywords:

Amiloride, 2H-chromene azo dye, Fluorescence quenching, Flow injection analysis

INTRODUCTION

compounds due to their premium physicochemical features such as Amiloride (AMD) [chemically it is known as [3,5-diamino-N-(diaminomethylene)-6-chloropyrazinecarboxamide] is a pyrazine optical properties [18, 19], stability [20], and their extensive applications in liquid crystals [21], chemosensors [22] and carbonyl guanidine derivative (fig. 1). It is an orally administrated with diuretic and mild antihypertensive properties. AMD was indicated for the inhibition of sodium-potassium exchange in kidneys by blocking the distal membrane to advance the loss of sodium and potassium

reabsorption [1, 2]. AMD is generally used nanotubes [23]. Chromene (Benzopyran) derivatives play a significant role in the generation of highly efficient fluorescent dyes for synthetic fiber and daylight fluorescent pigments [24, 25].

Fig. 2 shows 2H-chromene azo dye [26], namely (2-(4-nitrophenyl)-N-(4-(phenyldiazenyl)-2H-chromen-4-amine) containing the therapeutically at most in combinatiochromene nucleus as a new derivative of azo dye.

hydrochlorothiazide [3, 4]. The British certified name of the combination was co-amilozide, the

common use of this combination for the treatment of nephrogenic diabetes insipidus [5] and in nitroglycerin therapy [6, 7]. Several analytical techniques have been reported for Amiloride determination, including spectrophotometry [8-10], high-performance liquid chromatography (HPLC)[11, 12], Atomic Emission spectrometry [13] and polarography [14].

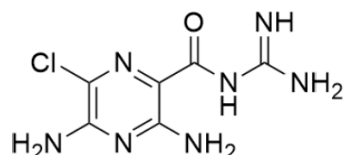
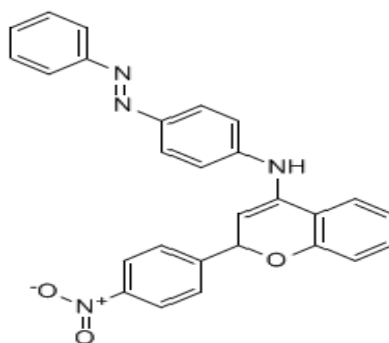


Fig. 1: Chemical structure of amiloride

Fig. 2: Chemical structure of the 2H-chromene azo dye

Azo compounds are the major kind of all synthetic



In the present work, a novel fluorometric method has been developed for the determination of Amiloride combined with a flow injection technique using a homemade ISNAG-fluorometer [27]. This method is based on using azo dye (2-(4-nitrophenyl)-N-(4- (phenyldiazenyl)-2H-chromen-4-amine) as fluorescence dye, then The flow the constant fluorescence is quenched measuring cell. The response of quenched fluorescence quenching was measured by ISNAG-fluorimeter.

MATERIALS AND METHODS

Apparatus and reagents

ISNAG-fluorometer is a homemade instrument [27] was used in fluorescence of azo dye was measured by a measuring the fluorescent response with 4-channels peristaltic pump (Ismatec, Switzerland), valve 6-port medium pressure injection valve (I D E X corporation, USA), sample loop (1 mm ID Optimization of variables chemical variable

dyes that are widely used in the world [15]. These compounds, with two phenyl rings detached by an azo (-N=N-) bond, are versatile molecules and have much importance in both academic and applied research [16, 17]. Azo dyes are an outstanding class of organic photoactive It was observed that 2H-chromene azo dye gives an emission band in ethanol at 425 nm after excitation with 387 nm (a maximum absorbance in ethanol), the fluorescence intensity of this dye was quenched after addition of the amiloride drug as fig. 3.

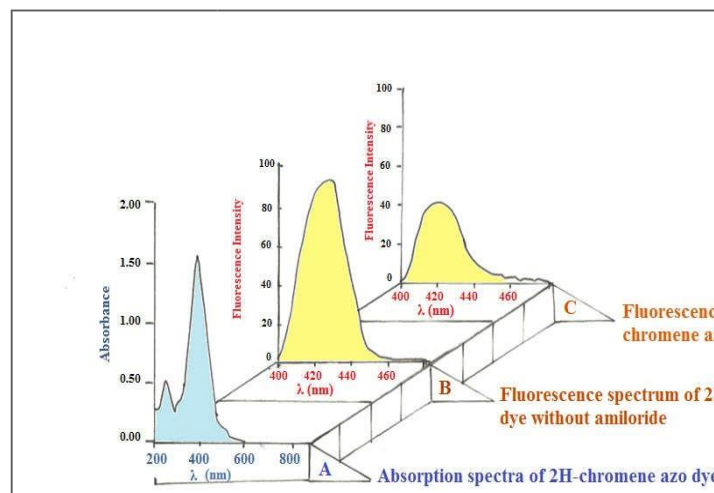


Fig. 3: (A) Absorption spectra of 2H-chromene azo dye in ethanol; fluorescence intensity of 2H-chromene azo dye in ethanol: (B) without amiloride (drug), (C) In the presence of drug

injection manifold system is composed of one line (fig. 4) represents the carrier stream (fluorescent azo dye solution (0.3 mmol/l) which passes through the injection valve to carry amiloride sample of 100 µl as a sample volume, 1.5 ml/min flow rate and 4 mmol/l concentration of amiloride) and then passes through the the continuous

homemade ISNAG- fluorimeter via low-pressure mercury lamp; it gives two main wavelengths, namely 184.9 nm and 253.7 nm. While the suggested probable mechanism pattern is expressed in scheme 1 [28, 29].

RESULTS AND DISCUSSION

using one line manifold system

Teflon, changeable length). The output signals were estimated via a Potentiometric recorder (Siemens, Germany (1-5 V)). UV-Vis Spectrophotometer digital double-beam mode (UV-Vis spectrophotometer, UV-1800, Shimadzu, Japan) was also used for classical spectrophotometric methods. SHIMADZU Fluorescence spectrometer to measure fluorescence spectra.

All chemicals were applied to analytical-reagent grade. A standard solution of AMD ($C_6H_8ClN_7O$, molecular weight 229.6 g/mol, 0.02 mol/l) was prepared by dissolving 1.148 g in 250 ml ethanol. 2H-

Effect of 2H-chromene azo dye concentration as a chemical variable

A set of 2H-chromene azo dye concentration ranging from 0.05-0.5 mmol/l were prepared as a carrier stream with a flow rate of 1.2 ml/min and 50 μ l of 4 mmol/l of amiloride as an injected sample to study the effect of the amiloride solution on the

quenching of continuous azo dye fluorescence. The profile shows in the fig. 5. A, B. Table 1 summed up the obtained results, which observed that an increase in fluorescence of azo dye with an increase of azo dye

chromene azo dye ($C_{27}H_{19}N_4O_3$, molecular synthesis previously [26], 0.001 mol/l).

Two pharmaceutical brands of amiloride concentration, at the same time, increased quenching effect by amiloride solution. The optimum concentration of azo dye that gave maximum fluorescence, quenching of continuous fluorescence by

purchased from a local pharmacy; (Actavis, UK) and (Moduretic, Lebanon).

amiloride, and minimum effect of blank response is 0.3 mmol/l. A higher concentration of azo dye does not include in this study due to the self-quenching [30].

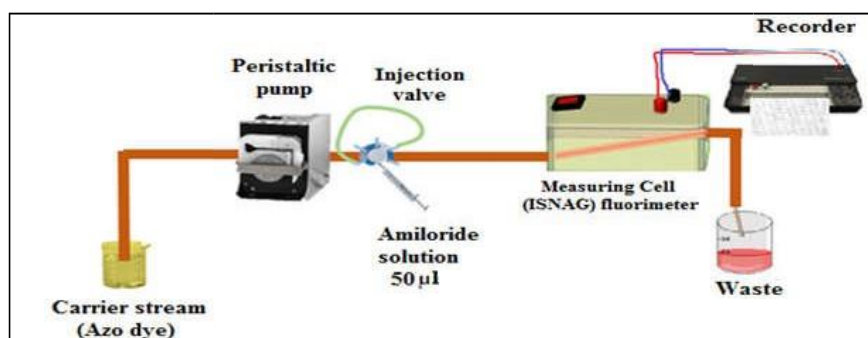
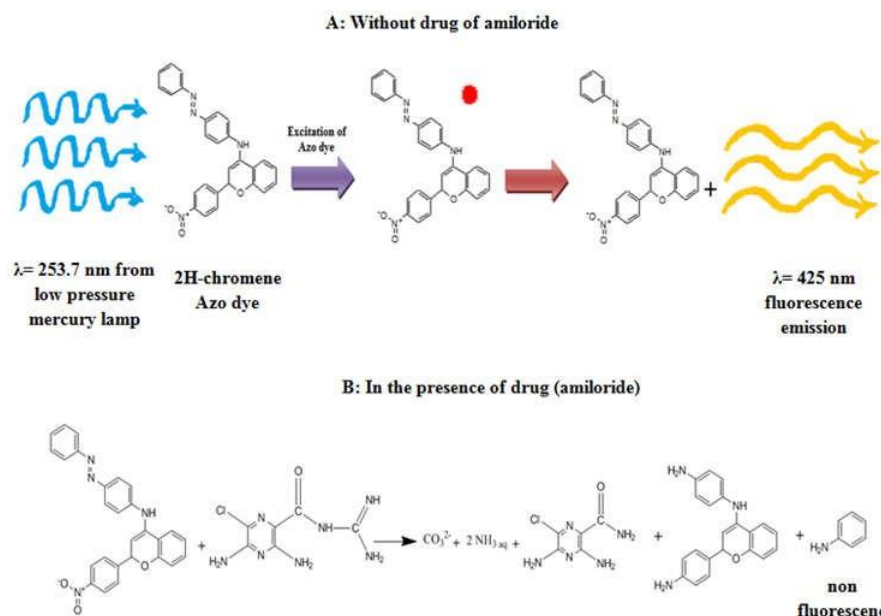


Fig. 4: Manifold system design of quenching of 2H-chromene azo dye fluorescence via the use of amiloride solution as an injected sample



Scheme 1: Proposed mechanism for quenching of 2H-chromene azo dye fluorescence

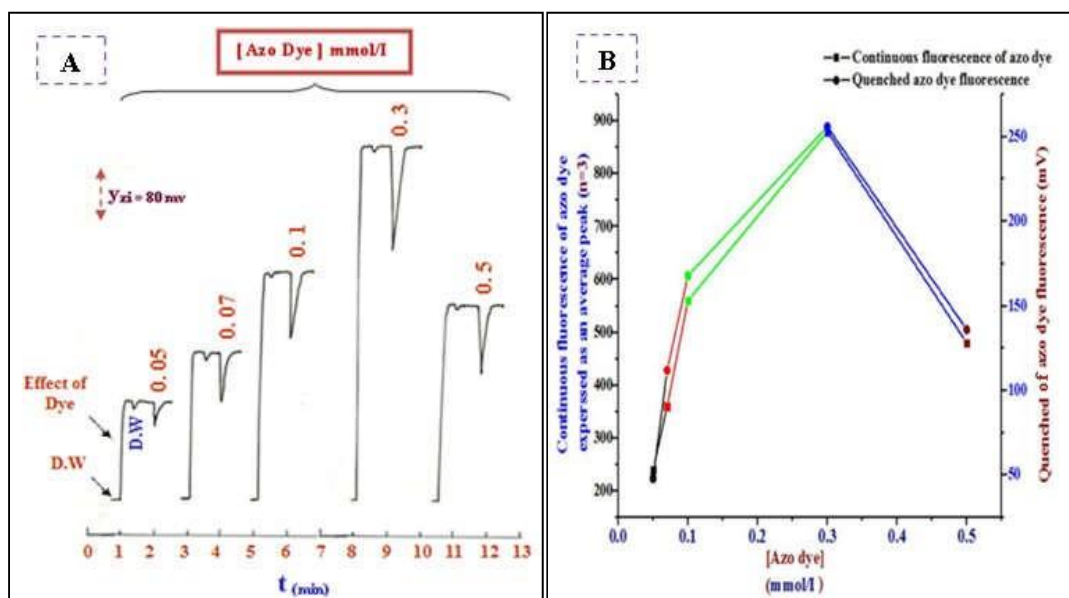


Fig. 5: Effect of 2H-chromene azo dye concentration on fluorescence intensity expressed as: (A) Profile versus time, (B) Increased of fluorescence using low-pressure mercury lamp characteristic emitted light at 184.9 and 253.7 nm, *DW Distilled water as a blank

Table 1: Effect of variation of azo dye concentration for determination of amiloride

[Azo dye] mmol/l	Continuous fluorescence of azo dye described as the average peak heights (n=3) $\bar{y}_i$ in mV	Quenched of azo dye fluorescence $\bar{y}_{Qi}$ (n=3) in mV	Confidence interval of the average response ^a (at 95% confidence level)
			$\bar{y}_i(\text{mV}) \pm t_{0.05/2, n-1} (\sigma_{n-1}/n)$
0.05	240	48	48 ± 1.23
0.07	360	112	112 ± 2.99
0.1	560	168	168 ± 2.12
0.3	880	256	256 ± 2.024
0.5	480	136	136 ± 2.34

Response of blank: 24mV, n= 3 (number of measurements), ^aData expressed as mean $\pm t_{0.05/2, n-1}$ (SEM), $t_{0.05/2, n-1} = 4.303$.

Physical parameters optimization Effect of flow rate

Employing the optimum concentration of azo dye (0.3 mmol/l) and a selected concentration 4 mmol/l of amiloride, 50 μl sample volume, and open valve mode with variable flow rate (0.5-2.0 ml/min) for the carrier stream (2H-chromene azo dye solution). Fig. 6A shows a kind of response profile that was obtained while all results were sorted in table 2. It can be observed that at a low flow rate, there is an expansion in dispersion and dilution effect due to the distribution of segment (i.e. amiloride) on a larger area [31]. But at the high flow rate result in an increase in peak height, the peak base width decrease, lower at analysis time, and minimize the dilution effect and obtained sharp edges responses [32]. Therefore, the best flow rate was 1.5 ml/min (fig. 6B).

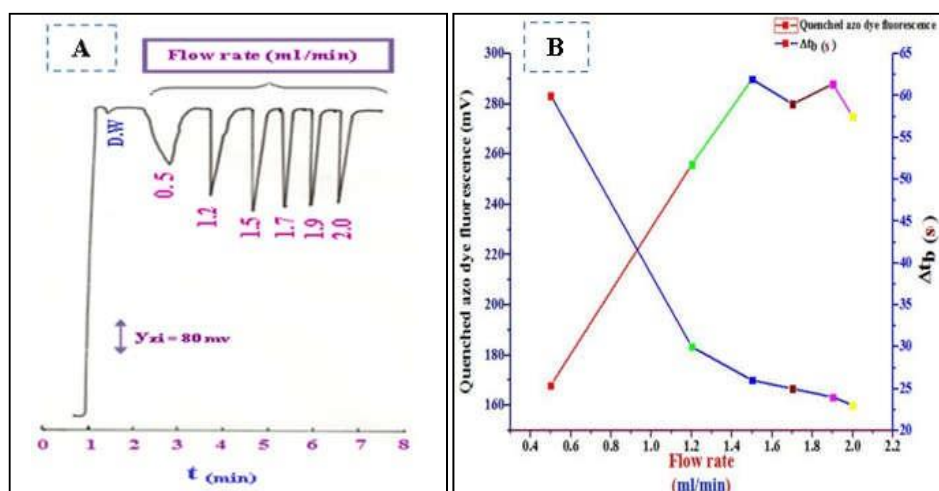


Fig. 6: Effect of the variance of flow rate on: (A) Response profile, (B) Quenching of fluorescence response by amiloride and the width of peak base (Δt_B)

Table 2: Effect of variable flow rate utilizing 50 μ l of 4 mmol/l of amiloride via quenching of 2H-chromene azo dye continuous fluorescence

Flow rate (ml/min)	Quenched of azo dye fluorescence (mV) depicted as the average peak heights (n=3) $\bar{y}_i$ (mV)	Confidence interval of the average response ^a (at 95% confidence level) ($\bar{y}_i \pm t_{0.05/2, n-1} (\sigma_{n-1} / \sqrt{n})$)	Δt_B Peak base width (s)
0.5	168	168 ± 1.57	60
1.2	256	256 ± 2.05	30
1.5	290	290 ± 2.11	26
1.7	280	280 ± 2.01	25
1.9	288	288 ± 2.03	24
2.00	275	275 ± 2.23	23

Continuous fluorescence response: 880mV, Response of blank: 24mV, n=3 (number of measurements), Δt_B (s): Time lapse for the quenched of azo dye fluorescence response within estimation cell or peak base width, ^aData expressed as mean $\pm t_{0.05/2, n-1}$ (SEM), $t_{0.05/2, n-1} = 4.303$.

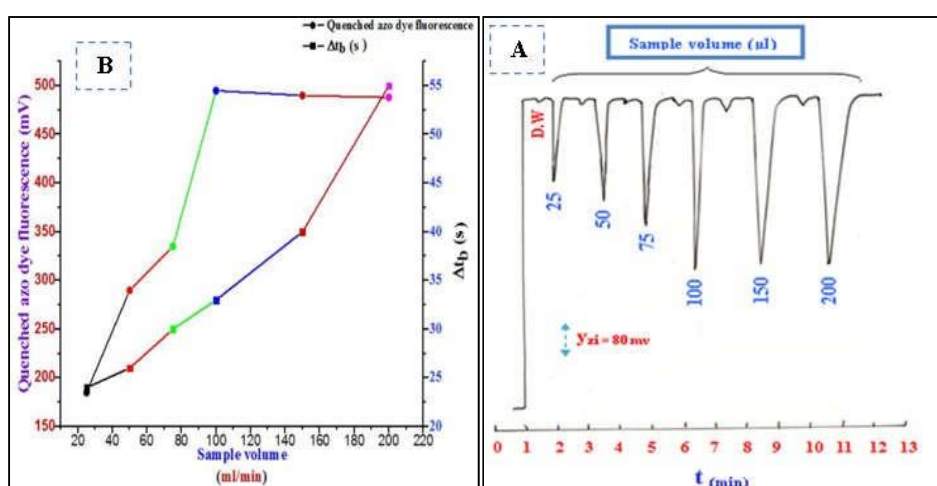


Fig. 7: Effect of variable sample volume on: (A) Profile using quenched fluorescence by amiloride, (B) Quenching of fluorescence response described as the average peak heights (mV) and Δt_B

Table 3: Effect of variable volume of sample on quenching of continuous fluorescence

Sample volume (μ l)	Quenched of azo dye fluorescence (n=3) mV expressed as the average peak heights (n=3) $\bar{y}_i$ (mV)	Confidence interval of the average response ^a (at 95% confidence level) $\bar{y}_i \pm t_{0.05/2, n-1} (\sigma_{n-1}/\sqrt{n})$	Δt_b Peak base width (s)
25	185	168 $\pm$ 1.57	24
50	290	256 $\pm$ 2.05	26
75	335	290 $\pm$ 2.11	30
100	495	280 $\pm$ 2.01	33
150	490	288 $\pm$ 2.03	40
200	488	275 $\pm$ 2.23	55

Response of continuous fluorescence: 880mV, n=3 (number of measurements), Δt_b (s): Time lapse for the Quenched of azo dye fluorescence response within measuring cell or peak base width, ^aData expressed as mean $\pm$ t_{0.05/2}, n-1 (SEM), t_{0.05/2}, n-1= 4.303.

Effect of variable sample volume

Adjusting all achieved optimum that was studied in previous sections with a variable volume of sample segment, which extends from 25-200 μ l in addition to open valve mode and 4 mmol/l amiloride concentration at flow rate 1.5 ml/min for carrier stream (0.3 mmol/l 2H-chromene azo dye solution) were used. All results are subjected in fig. 7. A, B and tabulated in table 3 which indicate clearly that the optimum sample volume is 100 μ l to obtain sharp and highest response profile expressed as a quenched of azo dye fluorescence, but an increase of the sample segment (>100 μ l) leads to a decrease in peak heights, which can be probably attributed to the long time period of the sample segment in front of a detector [33]. So that the 100 μ l was the most favorable choice to give the highest response for quenched continuous fluorescence of the azo dye molecule.

Calibration curve

A series of concentrations (0.03-10 mmol/l) of amiloride was prepared under the established optimum condition. Each measurement three times was repeated. The variation in quenched fluorescence response of ISNAG-fluorimeter with amyloid concentration was ranging from 0.03-8 mmol/l with linearity percentage ($R^2\%$): 98.79 % for dynamic range. Above 8 mmol/l; correlation coefficient value will deviate to the working range

at

linearity percentage ($R^2\%$): 97.59 % probably due to the quenching the inner fluorescence in the form of non-radiative thermal energy or internal convention between electronic levels of all fluorescent molecules and losing fluorescence energy and external convention. Fig. 8 shows the variation of response with concentration using linear regression treatment [34] for the newly developed methodology method. While the classical method using a spectrophotometer (the measurement of absorbance at 540 nm using Ce (IV)–Amiloride–H₃O system) with the range of (0.005-7) mmol/l [35].

The linear equation for dynamic range at n=13 is Quenched of fluorescence (n=3 ($\bar{Y}_{Qi}$)) in (mV) = 116.78 $\pm$ 24.13+84.99 $\pm$ 10.13 [amiloride] mmol. l⁻¹ While for working range at n= 14 is Quenched of fluorescence (n=3 ($\bar{Y}_{Qi}$)) in (mV) = 126.18 $\pm$ 33.19+79.32 $\pm$ 18.23 [amiloride] mmol. l⁻¹ and confidence level 95%.

The detection limit is calculated from the gradual dilution of the minimum concentration of the used of effective range calibration graph and depends on the values of a slope. Table 4 summarizes the different ways of calculating the limit of detection for the Quenched of fluorescence [34].

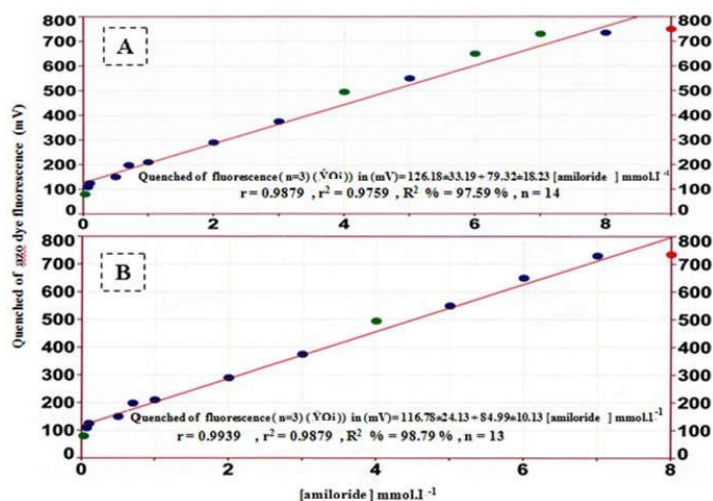


Fig. 8: Calibration graph of quenching 2H-chromene azo dye fluorescence response (mV) with amiloride concentrations (mmol/l): (A) range (0.03-10 mmol/l) and (B) range (0.03-8 mmol/l), n=13 or 14 (working range of calibration graph)

A repeated eight successive measurements of two concentrations (5 and 7 mmol/l) of analyte gave a variation of RSD % less than 0.5 % this indicates the method was distinguished by high accuracy with good repeatability [33]. Fig. 9 shows a kind of response profile.

Pharmaceutical drug analysis with ISNAG-fluorimeter

The performance of the newly established method (ISNAG- fluorimeter) was evaluated compared with the classical method (UV- vis spectrometer) by the determination of AMD in pharmaceutical drugs from various manufactures (amiloride, 5 mg- Actavis and Moduretic, 5 mg). The suggested method was applied to the analysis of standard addition, each Pharmaceutical drug by preparing a sequence of solutions from each drug via adding 0.2 ml (5 mmol/l) to five (10 ml) volumetric flasks. Then the standard solution (0.01 mmol/l) of AMD

is added in various amounts such as (0, 0.1, 0.2, 0.3 and 0.4 ml) and the flasks are diluted to the mark to get the concentration range between (0-0.4 mmol/l), table 5 summarizes the results obtained from the standard addition measurements which were mathematically treated at 95% confidence interval [34, 36].

Utilizing measurable chemometric treatment table 6 was obtained and the results of the two approaches were tried using paired t-test [27, 34]. The final deduction in table 6 displays that there is no significant difference between the newly proposed method and the classical method at confidence interval 95%, as the tabulated value is greater than the tcalculated value; Hence the recently developed method can be applied as a substitution mode for the determination of amiloride in pharmaceutical drugs.

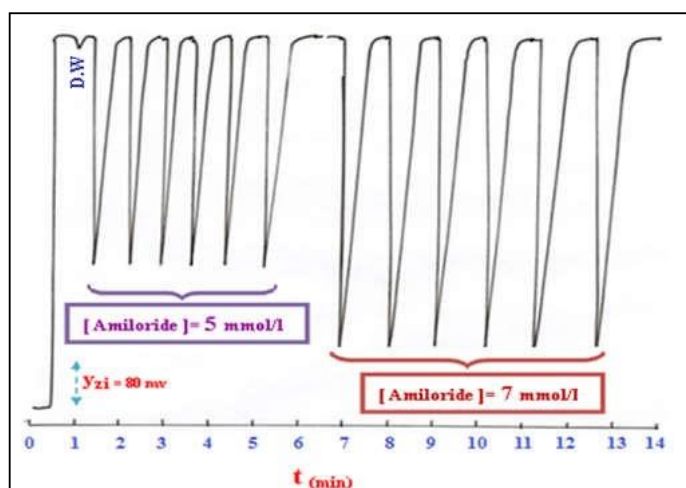


Fig. 9: Repeatability of six successive measurements of 5 and 7 mmol/l of amiloride via the quenching effect

CONCLUSION

In this work, a new method was developed and established for the determination of amiloride in pharmaceutical drugs using ISNAG- fluorimeter-CFI analysis and azo dye as a new reagent. The suggested method based on the quantitative quenching influence of amiloride drug on the native fluorescence of the azo dye. The statistical comparison via the t-test between this newly work and UV-Vis method was in good approval. Hence, this developed method can be used as an alternative method for the determination of amiloride in pharmaceutical drugs.

REFERENCES

José Molina, Pulgarrin JAM, and Lopez PF. Analysing amiloride levels in urine by use of isopotential fluorimetry. *Analyst*, 1997;122:247–52.

Authors: Bull MB and Laragh JH. Amiloride is a natriuretic that sparingly uses potassium. Published in 1968 in the journal *Circulation*, volume 37, pages 45–53.

Judson Chan, Alon U. For the purpose of treating congenital nephrogenic diabetic insipidus, hydrochlorothiazide and amiloride are prescribed. *United States Journal of Nephrology*, 1985, 5:9–13.

Clinical and basic pharmacology (Katzung BG). 10. McGraw-Hill Medical, 2006, USA: 10th ed. Kocher V, Seidl R, Kirchlechner V, and Waldhauser F. Nephrogenic diabetes insipidus treatment with amiloride and hydrochlorothiazide. *Child Health Arch* 1999;80:548–52.

*Potts JO, Farrell B, Parker AB, Parker JD. Diuretic treatment impacts exercise capacity and nitroglycerin tolerance in individuals with persistent stable angina. *Heart and Stroke* 1996;93:691-6.

Kent BA, McKay DW, Raju MK, and Campbell NR. While using diuretics, the antianginal effects of isosorbide dinitrate remain unchanged. *Therapeutics in Clinical Pharmacology*, 1994, 56: 229–34.

Ortiga An article authored by Barrales, Pellerano, Vazquez, and Molina Diaz. Direct solid-phase UV detection and cation exchange preconcentration allow for the quick and sensitive identification of amiloride. The article was published in *Anal Lett* in 2002 in number 35, pages 1491–504.

Saleh SS and Lotfy HM. Advances in UV spectrophotometry throughout the last ten years (2006–2016): a comprehensive overview. Published in the *International Journal of Pharmaceutical Science*, 2016;8:40–56.

Schuman TS, Ferraro MCF, Castellano PM. Chemometric analysis of pharmaceutical and synthetic combinations including amiloride hydrochloride, atenolol, hydrochlorothiazide, and timolol maleate. *Journal of Pharmaceutical and Biomedical Analytics*, 2004;34:305-14.

Mostafa A., El-Gindy A., and Emara S. The simultaneous measurement of atenolol, amiloride hydrochloride, and chlorthalidone was accomplished using HPLC and chemometric-assisted spectrophotometric techniques. *Pharmaceuticals* 2005;60:269–78.

Mohan JS, Rao NN. Pharmaceutical dosage form amiloride hydrochloride and furosemide were studied in a simultaneous manner utilising the reverse-phase high-performance liquid chromatography technique for both estimation and forced degradation. *Journal of Asian Pharmaceutical and Clinical Research* 2018;11:215–21.

Written by Issa YM, Shoukry AF, Ibrahim H, and Mohamed SK. By forming ion associations with ammonium reineckate, antazoline, hydralazine, amiloride, thiamine, and quinine may be determined using atomic emission spectrometry. Published in *Anal Lett* in 1994, volume 27, pages 731–734.

Authors: Jimenez AI, Arias JJ, Jimenez F., Hernandez OM, Martin ME. Partial least-squares approach in analysis via differential pulse polarography. Medicinal formulations containing amiloride and hydrochlorothiazide were determined simultaneously. In 1999, the *Anal Chim Acta* published an article with the DOI number 381:247–56.

Collaborators: Nisal A, Trivedy K, Mohammad H,

Panneri S, Gupta S, Lele A, and others. Using a green feeding technique, silk glands absorb azo dyes to create coloured silk cocoons. The article is published in ACS Sustainable Chem Eng and has the online journal article number 2:312–314. Adosraku RK, Kofie W, Dzidzoramengor C. Making azo dyes and testing their antibacterial capabilities. The article "Int J Pharm Pharm Sci 2014;7:398-401" provides further reference information.

J. Sahoo and S. Paidesetty. A overview of the medicinal potential of organic molecules based on azo groups. Journal of Asian Pharmaceutical and Clinical Research, 2016;9 Suppl 1:33-9. Joshua J. An innovative anion sensor that combines fluorescence and colorimetry; it uses a coumarin group as its signal unit and a urea group as its binding site. Pigments and Dyeing, 2010;87:272-6.

Whitehead OA, Coe BJ. Dinuclear cyclopalladated complexes of push-pull azobenzenes: syntheses, electronic structures, and dichroic behaviour. Chapter 30: Organometallics, 2011: 2212-22. Ye Yong and Ikeda T. Liquid crystals and smart materials that respond to light include azobenzene. 2009, 1st edition. Wiley, Hoboken (NJ).

Pavel A. Bogdanov and Alexander V. Vorobiev. Using domain structure rearrangement, photo-orientation of liquid-crystalline materials containing azobenzene is achieved. The Citation is from the Journal of Physical Chemistry B, Volume 117, Pages 13936–13945, 2013. This work was authored by Li T, Yang Z, Li Y, Liu Z, Qi G, and Wang B. A new kind of fluorescein to be used as a colorimetric chemosensor to detect copper (II) ions. Pigment Dyeing 2011;88:103–08.

Yu L., Banerjee IA., and Matsui H. Molecular recognition enables the photochemically controlled immobilisation of azobenzene nanotubes on patterned α -CD monolayer/au substrates, using host-guest chemistry in the construction of nanotube-based devices. The citation for this article is as follows: J Am Chem Soc 2003;125:9542-3.

Bahra GS, Millar V, and Griffiths J. The effect of electron acceptor residues and chain length on 3-substituted 7-N, N-diethylaminocoumarin dyes. Pigment Dyes 1995;28:327–39. Minami M, Machida M, Oda K, Ohno K, Nishizono N. The luminous characteristics and reactivity with isatin of 3-Aryl-7 diethylaminocoumarin derivatives synthesised using chemical synthesis. "Heterocycles" (2001, 55: 1897-906). Published by Nafea HM and Al-Kawaz AMN. Metal ion (II) coordinates with 2h-chromene azo

novel ligands: synthesis, characterisation, antibacterial, DNA cleavage, and fluorescence activities. Worldwide Journal of Pharmaceutical Research and Science, 2020, 11, 1953–1960. Abd-Alrazack HF and Turkie NS. The use of a handmade ISNAG fluorimeter in conjunction with CFIA is a newly established approach for the determination of indomethacin using potassium hexacyanoferrate (III). The article is published in the Journal of Global Pharmacology and Technology in 2018 and has the the DOI: 10.447-60.

Spectroscopy by Lakowicz JR. Third edition. New York: Springer; 2006.

First edition of Smith's Organic Chemistry, published in 2006 by MC Graw-Hill in New York.